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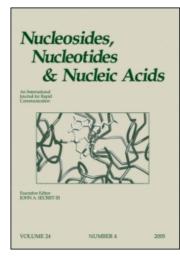
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Synthesis and Biological Activity of 2-Aminopurine Methylenecyclopropane Analogues of Nucleosides

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ABSTRACT

Synthesis and biological activity of racemic 2-aminopurine methylenecyclopropane analogues of nucleosides 4, 5, 10 and 11 is described. One-pot alkylationelimination of 2-aminopurine (6) with dibromide 7 gave a mixture of four isomeric methylenecyclopropanes. The (E, Z)- N^9 and (E, Z)- N^7 isomers 8 and 9 were resolved by chromatography on silica gel. Deacetylation of 8 afforded the respective (Z)-N⁹ and (E)-N⁹ isomers **4** and **10** which were separated by chromatography on silica gel. In a similar fashion, (E, Z)-N⁷ mixture 9 furnished (Z)-N⁷ and (E)-N⁷ isomers **5** and **11**. The S-(+)-enantiomer **4** was obtained by desulfurization of (S)-(+)-6-thiosynguanol (13) with Raney Ni. Compound 13 was obtained from (S)-(+)-2-amino-6-chloro derivative 12 and NaSH in methanol. Racemic analogues 4, 5, 10 and 11 were inactive against HCMV, HSV-1,

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HSV-2, EBV and VZV. Enantiomer (S)-(+)-4 inhibited replication of HSV-1 in BSC-1 cells (ELISA) with EC₅₀ 35 μ M and it was non-cytotoxic in KB cells (CC₅₀ > 100 μ M). Compound (S)-(+)-4 was also moderately effective against VZV in HFF culture with EC₅₀/CC₅₀ (μ M) 60/ > 460 and it was a substrate for xanthine oxidase.

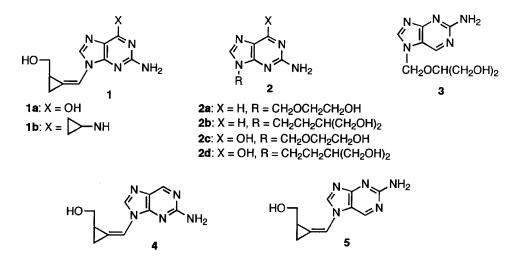
Key Words: Nucleoside analogues; Methylenecyclopropanes; 2-aminopurine; Prodrugs; Alkylation-elimination; Xanthine oxidase.

INTRODUCTION

Purine Z-methylenecyclopropane analogues of nucleosides are broad-spectrum antiviral agents strongly effective against human cytomegalovirus (HCMV). The anti-CMV activity is associated with the S-(+) enantiomeric forms of these analogues. Synguanol (1a) and its cyclopropylamino analogue 1b are effective in vivo in various mouse models of CMV infection. Efficacy of compound 1b, a potential prodrug of synguanol (1a), is equal or superior to current drug ganciclovir.

The 6-deoxyanalogues of acyclovir and penciclovir (**2a** and **2b**) are efficient inhibitors of herpesviruses in vivo although they exhibit little activity in cell culture. Both analogues act in vivo as prodrugs that are converted to guanine analogues **2c** and **2d** by the action of xanthine oxidase. More recently, the N⁷-isomer of 6-deoxyganciclovir **3** was found to exhibit a potent activity against herpesviruses in vitro and in vivo. [8–11] Unlike the N⁹-isomers **2a** and **2b**, analogue **3** does not require oxidation in vivo for activity.

Therefore, it was of interest to synthesize and investigate the N^9 - and N^7 -isomers of methylenecyclopropane analogues 4 and 5.



RESULTS AND DISCUSSION

A. Synthesis

The N⁷- and N⁹-isomers of 2-aminopurine nucleoside analogues are usually synthesized from the corresponding guanine derivatives. [12] For our purpose, it was considered advantageous to prepare both isomers **4** and **5** directly from commercially available 2-aminopurine (**6**) in a one-pot reaction (Sch. 1) by alkylation-elimination procedure employed previously with other nucleic acid bases. [1,13,14] Reaction of 2-aminopurine (**6**) with dibromide **7** and Cs₂CO₃ in DMF at 65°C and chromatographic separation gave (E, E)-N⁹-isomers **8** (53%) and (E, E)-N⁷-isomers **9** (20%). The N⁹/N⁷ ratio (2.7/1) is significantly lower than that observed in simple alkylations of 2-aminopurine (**6**) (5/1 – 11/1). [15] Deprotection of **8** using K₂CO₃ in aqueous methanol gave, after chromatographic separation, compound **4** (43%) and the E-isomer **10** (33%). A similar treatment of **9** afforded analogue **5** (49%) and the E-isomer **11** (35%).

The S-(+)-enantiomer of **4** was prepared as follows (Sch. 2). The (S)-(+)-2-amino-6-chloro derivative $(12)^{[14]}$ was converted to 6-thiosynguanol (13) by reaction with NaSH in methanol in 67% yield. Desulfurization of **13** with Raney Ni in aqueous methanol gave S-(+)-**4** (46%). The isomeric assignments were confirmed by UV and NMR spectroscopy (Table 1). As expected, [12] the long wavelength UV maximum of the N⁷-isomers **5** and **11** is bathochromically shifted relative to N⁹-isomers **4** and **10**. Also, the patterns of chemical shifts δH_8 (N⁹) $< \delta H_8$ (N⁷), δNH_2 (N⁹) $> \delta NH_2$ (N⁷) and C₈ (N⁹) $< C_8$ (N⁷) follow an empirical rule postulated for N⁹- and N⁷-alkyl purines. [16] The Z, E-isomeric assignment of N⁹-isomers then

Scheme 1.



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Scheme 2.

agreed with those of other purine methylenecyclopropane analogues:^[1] Deshielding of the H_8 , OH and shielding of the $H_{1'}$ in the Z-isomer 4 relative to the E-isomer 10. Non-equivalency of the H_5 protons in the Z-isomer 4 as opposed to E-isomer 10 is also an important feature found generally in the purine series.^[1,2] Interestingly, these relationships were closely followed by N^7 isomers 5 and 11. It should be noted that in an *anti* conformation of both Z-isomers 4 and 5 the CH₂OH is juxtaposed to the H_8 which explains a significant downfield shift of the latter resonance relative to the respective E-isomers 10 and 11.

B. Biological Activity

Compounds **4**, **5**, **10** and **11** were inactive (EC₅₀ > 100 μ M) in the following assays: HCMV, HSV-1, HSV-2, EBV, HBV and VZV. Enantiomer *S*-(+)-**4** was moderately effective against HSV-1/BSC-1 (ELISA), cytotoxicity in KB cells, EC₅₀/CC₅₀ (μ M) 35/ > 100 and VZV/HFF /EC₅₀/CC₅₀ (μ M) 35/ > 100.

Compound (S)-(+)-**4** was a substrate for xanthine oxidase (Figs. 1 and 2). The reaction lacks stereoselectivity because racemic Z- and E-isomers **4** and **10** were also oxidized (data not shown). The assays were performed under conditions close to those

Table 1. UV maxima and some relevant NMR chemical shifts of compounds 4, 5, 10, and 11.

Compound	λ_{max} (nm)	δH_8	δH_{1^\prime}	δОН	δNH_2	δH_{5^\prime}	C ₈
4 , Z-N ⁹ 5 , Z-N ⁷ 10 , E-N ⁹	310 330 310	8.60 8.81 8.45	7.25 7.51 7.35	5.07 5.08 4.80	6.63 6.35 6.78	3.73 & 3.28 3.63 & 3.31 3.40	140.1 145.6 139.6
11, E -N ⁷	330	8.57	7.58	4.91	6.37	3.41	146.4

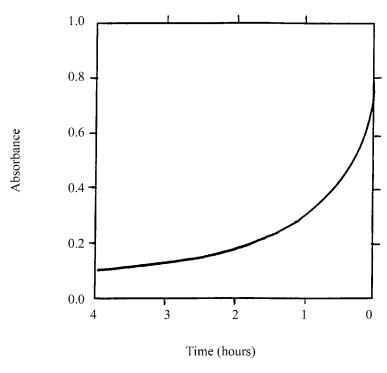


Figure 1. Time-course of oxidation of (S)-(+)-6-deoxysynguanol (4) to (S)-(+)-synguanol [(S)-(+)-1a] catalyzed by xanthine oxidase. *Conditions*: 0.07 mM (S)-(+)-4 in 0.05 M Na₂HPO₄ (pH 7.4, 1.2 mL) plus xanthine oxidase from cow milk (20 units/mL, 20 μ L, 0.4 units), room temperature, continuous monitoring at 304 nm.

reported for 6-deoxypenciclovir (2b).^[7] These results have indicated that (S)-(+)-6-deoxysynguanol (4) is a viable candidate for in vivo testing as a prodrug of (S)-(+)-synguanol (1a).

EXPERIMENTAL SECTION

General Methods

The ¹H and ¹³C NMR spectra were determined at 400 and 100 MHz, respectively, in CD₃SOCD₃ unless stated otherwise. UV spectra were measured in ethanol. Mass spectra were performed in an electrospray ionization mode (ESI-MS) using aqueous MeOH and NaCl.

(Z,E)-2-Amino-9-[(2-acetoxymethyl)cyclopropylidenemethyl]purine (8) and (Z,E)-2-Amino-7-[(2-acetoxymethyl)cyclopropylidenemethyl]purine (9). A mixture of 2-aminopurine (6, 600 mg, 4.44 mmol) and dibromide 7 (1.52 g, 5.33 mmol) in DMF (30 mL) was stirred with Cs_2CO_3 (7.24 g, 22.2 mmol) at room temperature for 2 h and then at 65°C for 6 h. The solids were filtered off, the filtrate was evaporated in



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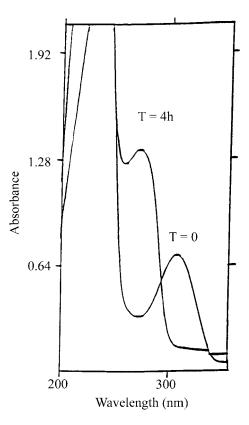


Figure 2. UV spectra of (S)-(+)-4 (pH 7.4, T=0, before addition of enzyme) and after incubation with enzyme (T=4 h). The latter spectrum corresponded to authentic (S)-(+)-synguanol [(S)-(+)-1a].

vacuo and the residue was chromatographed on a silica gel column using CH_2Cl_2 : MeOH = 98:2) as an eluent. The (E, Z)-9-isomers **8** (610 mg, 53%) were eluted first followed by (E, Z)-7-isomers **9** (230 mg, 20%).

(*E, Z*)-9-isomers **8**: Mp 120–124°C; UV max 310 nm (ϵ 6,000), 232 (ϵ 24,600); ¹H NMR (CDCl₃) δ 8.70 and 8.68 (s, 1H), 8.29 and 8.21 (s, 1H), 7.48 and 7.36 (dd, 1H, J=4.0 and 2.4 Hz), 5.30 (brs, 2H), 4.33–3.95 (m, 2H), 2.29–2.13 (m, 1H), 2.10 and 2.05 (s, 3H), 1.80–1.76 and 1.68–1.63 (m, 1H), 1.47–1.43 and 1.36–1.32 (m, 1H); ¹³C NMR 171.3, 171.1, 160.4, 160.3, 152.0, 150.2, 150.1, 140.0, 139.1, 128.2, 113.9, 113.1, 111.6, 111.5, 67.0, 66.3, 21.2, 21.1, 16.4, 14.4, 10.3, 8.0; ESI-MS 260 (M+H, 12.0), 282 (M+Na, 100.0).

(*E, Z*)-7-isomers 9: Mp 176–180°C; UV max 326 nm (ϵ 4,800), 250 (ϵ 14,100), 211 (ϵ 22,600); ${}^{1}H$ NMR δ 8.83 and 8.76 (s, 1H), 8.53 and 8.51 (s, 1H), 7.62 and 7.53 (d, 1H, J=1.6 Hz), 6.30 (brs, 2H), 4.16–3.80 (m, 2H), 2.43–2.39 and 2.13–2.09 (m, 1H), 2.03 and 1.77 (s, 3H), 1.92–1.88 and 1.66–1.62 (m, 1H), 1.64–1.60 and 1.37–1.33 (m, 1H); ${}^{13}C$ NMR (CDCl₃) 170.9, 170.5, 163.1, 163.0, 161.8, 146.5, 146.0, 143.4, 143.2, 118.1, 115.0, 114.5, 114.3, 113.3, 66.6, 66.2, 21.4, 21.0, 16.7, 14.8, 11.6, 8.2; ESI-MS 260 (M+H, 10.8), 282 (M+Na, 100.0).

(*Z*)-2-Amino-9-[(2-hydroxymethyl)cyclopropylidenemethyl]purine (4) and (*E*)-2-Amino-9-[(2-hydroxymethyl)cyclopropylidenemethyl]purine (10). The isomeric mixture 8 (520 mg, 2.0 mmol) was stirred with K_2CO_3 (304 mg, 2.2 mmol) in MeOH- H_2O (15:1, 30 mL) at room temperature for 3 h. The pH was adjusted to 7 with AcOH, volatile components were evaporated and the residue was chromatographed on a silica gel column using CH_2Cl_2 :MeOH = 19:1 as an eluent to give the *Z*-isomer 4 (185 mg, 43%) followed by *E*-isomer 10 (145 mg, 33%) as white solids.

Z-isomer 4: Mp 186–188°C; UV max 310 nm (ε 7,800), 233 (ε 33,100); 1 H NMR δ 8.68 (s, 1H, H₆), 8.60 (s, 1H, H₈), 7.25 (d, 1H, J=1.6 Hz, H₁), 6.63 (s, 2H, NH₂), 5.07 (dd, 1H, J=6.2 and 5.2 Hz, OH), 3.73 (m, 1H) and 3.28 (m, overlapped with H₂O, H₅), 2.11 (m, 1H, H₄), 1.48 (dt, 1H) and 1.20 (m, 1H, H₃); 13 C NMR 161.5 (C₂), 152.0 (C₄), 150.1 (C₆), 140.1 (C₈), 127.2 (C₅), 115.7 (C₂), 110.3 (C₁), 63.6 (C₅), 19.9 (C₄), 6.9 (C₃); ESI-MS 218 (M+H, 95.8), 240 (M+Na, 100.0). Anal. Calcd for C₁₀H₁₁N₅O·0.5H₂O: C, 53.09; H, 5.35; N, 30.96. Found: C, 53.38; H, 5.40; N, 31.12.

E-isomer 10: Mp 124–126°C; UV max 310 nm (ε 7,500), 233 (ε 30,600); ¹H NMR δ 8.65 (s, 1H, H₆), 8.45 (s, 1H, H₈), 7.35 (dd, 1H, J = 4.2 and 2.4 Hz, H_{1'}), 6.78 (s, 2H, NH₂), 4.80 (brs, 1H, OH), 3.40 (d, overlapped with H₂O, J = 6.8 Hz, H_{5'}), 1.96 (m, 1H, H_{4'}), 1.70 (dt, 1H) and 1.36 (m, 1H, H_{3'}); ¹³C NMR 161.5 (C₂), 152.2 (C₄), 150.2 (C₆), 139.6 (C₈), 127.2 (C₅), 116.4 (C_{2'}), 110.5 (C_{1'}), 63.9 (C_{5'}), 18.3 (C_{4'}), 10.0 (C_{3'}); ESI-MS 218 (M+H, 100.0), 240 (M+Na, 66.8). Anal. Calcd for $C_{10}H_{11}N_5O$: C, 55.29; H, 5.10; N, 32.24. Found: C, 55.13; H, 5.19; N, 32.13.

(*Z*)-2-Amino-7-[(2-hydroxymethyl)cyclopropylidenemethyl]purine (5) and (*E*)-2-Amino-7-[(2-hydroxymethyl)cyclopropylidenemethyl]purine (11). A mixture of 7-isomers 9 (230 mg, 0.88 mmol) was deacetylated as described for acetates 8 to give the *Z*-isomer 5 (95 mg, 49%) followed by *E*-isomer 11 (65 mg, 35%) as white solids.

Z-isomer **5**: Mp 211–213°C; UV max 330 nm (ϵ 5,100), 253 (ϵ 16,400), 210 (ϵ 22,700); ¹H NMR δ 8.84 (s, 1H, H₆), 8.81 (s, 1H, H₈), 7.51 (d, 1H, J=1.6 Hz, H₁'), 6.35 (s, 2H, NH₂), 5.08 (brs, 1H, OH), 3.63 (dd, 1H, J=11.2, 6.4 Hz) and 3.31 (dd, 1H, J=11.4, 7.6 Hz, H₅'), 2.19 (m, 1H, H₄'), 1.49 (poorly resolved dt, 1H) and 1.20 (m, 1H, H₃'); ¹³C NMR 162.8 (C₂), 161.8 (C₄), 145.6 (C₈), 143.1 (C₆), 118.2 (C₅), 115.4 (C₂'), 113.2 (C₁'), 63.5 (C₅'), 20.4 (C₄'), 7.1 (C₃'); ESI-MS 218 (M+H, 89.8), 240 (M+Na, 78.4), 457 (2M+Na, 100.0). Anal. Calcd for C₁₀H₁₁N₅O: C, 55.29; H, 5.10; N, 32.24. Found: C, 55.41; H, 5.16; N, 32.23.

E-isomer 11: Mp 202–204°C; UV max 330 nm (ε 5,100), 254 (ε 16,200), 212 (ε 19,300); 1 H NMR δ 8.85 (s, 1H, H₆), 8.57 (s, 1H, H₈), 7.58 (s, 1H, H_{1'}), 6.37 (s, 2H, NH₂), 4.91 (brs, 1H, OH), 3.41 (m, 2H, H_{5'}), 1.96 (m, 1H, H_{4'}), 1.78 (poorly resolved dt, 1H) and 1.46 (m, 1H, H_{3'}); 13 C NMR 163.0 (C₂), 161.8 (C₄), 146.4 (C₈), 143.5 (C₆), 118.1 (C₅), 115.4 (C_{2'}), 113.9 (C_{1'}), 63.8 (C_{5'}), 18.6 (C_{4'}), 10.9 (C_{3'}); ESI-MS 218 (M+H, 9.0), 240 (M+Na, 77.3), 457 (2M+Na, 100.0). Anal. Calcd for C₁₀H₁₁N₅O: C, 55.29; H, 5.10; N 32.24. Found: C, 55.42; H, 5.12; N 32.03.

(S, Z)-(+)-2-Amino-6-thio-9-[(2-hydroxymethyl)cyclopropylidenemethyl]purine (13). A mixture of compound 12 (400 mg, 1.6 mmol) and sodium hydrosulfide hydrate (3.00 g, 53.5 mmol) in methanol (50 mL) was stirred at room temperature under N_2 for 24 h. The solvent was evaporated, the residue was dissolved in water

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(40 mL) and pH of the solution was adjusted to 6 with acetic acid. The precipitate was filtered off and it was washed with water. Recrystallization from methanol-water (1:1) gave compound **13** (265 mg, 67%). Mp. 240°C (decomp.); [α]_D²⁰ 91.0° (c 0.26, DMF); UV max 346 nm (ε 24,000), 237 nm (ε 26,600); ¹H NMR δ 1.19 (ddd, 1H), 1.45–1.49 (dt, 1 H), 2.11 (m, 1H), 3.28 (m, overlapped with H₂O), 3.70 (m, 1H), 5.05 (t, 1H, J=5.5 Hz), 6.85 (s, 2H), 7.09 (d, 1H, J=2.0 Hz), 8.48 (s, 1H), 11.96 (s, 1H); ¹³C NMR 6.9, 19.8, 63.4, 110.5, 116.8, 128.6, 137.9, 146.8, 153.9, 175.7. ESI-MS 521 (2M + Na, 50), 272 (M + Na, 100), 250 (M + H, 10). Anal. Calcd for $C_{10}H_{11}N_5OS$: C, 48.18; H, 4.45; N, 28.09; S, 12.86. Found: C, 48.10; H, 4.61; N, 28.25; S, 13.02.

(*S*, *Z*)-(+)-2-Amino-9-[(2-hydroxymethyl)cyclopropylidenemethyl]purine [(*S*)-(+)-4]. Raney Ni (200 mg) was added to a solution of compound 13 (166 mg, 0.67 mmol) in water: MeOH (1:1, 100 mL). The reaction mixture was stirred at room temperature for 16 h, catalyst was filtered off and filtrate was evaporated. The crude product was chromatographed on a silica gel column CH₂Cl₂: MeOH (9:1) to give compound (*S*)-(+)-4 (66 mg, 46%). Mp 185–187°C; [α]_D²⁰ 77.7° (c 0.30, DMF); UV 310 nm (ϵ 7,900), 232 (ϵ 32,400); ¹H and ¹³C NMR were identical with those of racemic compound 4; ESI-MS 457 (2 M + Na, 21.2), 240 (M + Na, 100.0), 218 (M + H, 17.9). Anal. Calcd for C₁₀H₁₁N₅O: C, 55.29; H, 5.10; N, 32.24. Found: C, 55.40; H, 5.10; N, 32.19.

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